




ERS technical standard on bronchial challenge testing: pathophysiology and methodology of indirect airway challenge testing

Teal S. Hallstrand¹, Joerg D. Leuppi², Guy Joos³, Graham L. Hall⁴, Kai-Håkon Carlsen⁵, David A. Kaminsky⁶, Allan L. Coates⁷, Donald W. Cockcroft⁸, Bruce H. Culver¹, Zuzana Diamant^{9,10}, Gail M. Gauvreau¹¹, Ildiko Horvath¹², Frans H.C. de Jongh¹³, Beth L. Laube¹⁴, Peter J. Sterk¹⁵ and Jack Wanger¹⁶, the American Thoracic Society (ATS)/ European Respiratory Society (ERS) Bronchoprovocation Testing Task Force

 @ERSpublications
This international task force recently reported the general considerations for “direct” bronchoprovocation testing. In the present document, the task force provides updated recommendations on the pathophysiology and conduct of “indirect” challenge tests. <http://ow.ly/FR1K30m99Ef>

Cite this article as: Hallstrand TS, Leuppi JD, Joos G, *et al.* ERS technical standard on bronchial challenge testing: pathophysiology and methodology of indirect airway challenge testing. *Eur Respir J* 2018; 52: 1801033 [<https://doi.org/10.1183/13993003.01033-2018>].

ABSTRACT Recently, this international task force reported the general considerations for bronchial challenge testing and the performance of the methacholine challenge test, a “direct” airway challenge test. Here, the task force provides an updated description of the pathophysiology and the methods to conduct indirect challenge tests. Because indirect challenge tests trigger airway narrowing through the activation of endogenous pathways that are involved in asthma, indirect challenge tests tend to be specific for asthma and reveal much about the biology of asthma, but may be less sensitive than direct tests for the detection of airway hyperresponsiveness. We provide recommendations for the conduct and interpretation of hyperpnoea challenge tests such as dry air exercise challenge and eucapnic voluntary hyperpnoea that provide a single strong stimulus for airway narrowing. This technical standard expands the recommendations to additional indirect tests such as hypertonic saline, mannitol and adenosine challenge that are incremental tests, but still retain characteristics of other indirect challenges. Assessment of airway hyperresponsiveness, with direct and indirect tests, are valuable tools to understand and to monitor airway function and to characterise the underlying asthma phenotype to guide therapy. The tests should be interpreted within the context of the clinical features of asthma.

This document was endorsed by the European Respiratory Society on September 19, 2018, and by the American Thoracic Society on August 27, 2018.

Received: June 01 2018 | Accepted after revision: July 20 2018

Copyright ©ERS 2018

Affiliations: ¹Division of Pulmonary, Critical Care and Sleep Medicine, University of Washington, Seattle, WA, USA. ²University Clinic of Medicine, Cantonal Hospital Baselland, Liestal, and Medical Faculty University of Basel, Basel, Switzerland. ³Dept of Respiratory Medicine, University of Ghent, Ghent, Belgium. ⁴Children's Lung Health, Telethon Kids Institute, School of Physiotherapy and Exercise Science, Curtin University, and Centre for Child Health Research University of Western Australia, Perth, Australia. ⁵University of Oslo, Institute of Clinical Medicine, and Oslo University Hospital, Division of Child and Adolescent Medicine, Oslo, Norway. ⁶Pulmonary and Critical Care, University of Vermont College of Medicine, Burlington, VT, USA. ⁷Division of Respiratory Medicine, Translational Medicine, Research Institute-Hospital for Sick Children, University of Toronto, ON, Canada. ⁸Division of Respiratory, Critical Care and Sleep Medicine, Royal University Hospital, Saskatoon, SK, Canada. ⁹Dept of Clinical Pharmacy and Pharmacology and QPS-Netherlands, University Medical Centre Groningen, University of Groningen, Groningen, The Netherlands. ¹⁰Dept of Respiratory Medicine and Allergology, Lund University, Lund, Sweden. ¹¹Division of Respiratory, Dept of Medicine, McMaster University, Hamilton, ON, Canada. ¹²Dept of Pulmonology, National Korányi Institute of Pulmonology, Budapest, Hungary. ¹³Dept of Pulmonary Medicine, Medisch Spectrum Twente, Enschede, The Netherlands. ¹⁴Division of Pediatric Pulmonology, The Johns Hopkins University School of Medicine, Baltimore, MD, USA. ¹⁵Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands. ¹⁶Pulmonary Function Testing and Clinical Trials Consultant, Rochester, MN, USA.

Correspondence: Teal S. Hallstrand, Division of Pulmonary, Critical Care and Sleep Medicine, Center for Lung Biology, University of Washington, Box 358052, Seattle, WA 98109-4714, USA. E-mail: tealh@u.washington.edu

Introduction

Bronchoprovocation tests measure the propensity to develop airflow obstruction when the airways are challenged with either a direct-acting stimulus such as methacholine, or indirectly by physical or pharmacological stimuli that induce airway narrowing through the activation of inflammatory or neuronal cells (figure 1). The measurement of such indirect airway hyperresponsiveness (AHR) is valuable as a diagnostic test for asthma, and can be used to understand the underlying pathophysiology of asthma and as a guide for therapy. In the first technical standard on bronchial challenge testing, we described the general considerations for bronchoprovocation testing and updated the method to perform a direct bronchoprovocation challenge test with aerosolised methacholine [1]. Here, we describe the basis for the indirect challenge tests and the most commonly used methods to conduct these tests.

In a broad sense, direct challenge tests such as methacholine are sensitive to detect asthma, but are not entirely specific for asthma, as subjects with other airway disorders may have direct AHR. A negative methacholine challenge test in a currently symptomatic patient is highly suggestive of a diagnosis other than asthma; however, there are rare instances where indirect AHR is present in the absence of direct AHR. In contrast, indirect challenge tests act through the generation of mediators from cells present in the airways, providing an assessment of the mechanisms that are involved in asthma pathophysiology. Because of this basis, indirect tests tend to be more specific for asthma, but generally less sensitive than direct tests to detect AHR and support diagnosis of asthma in the appropriate clinical context (figure 2) [2–4]. Thus, the indirect challenge tests are useful to understand the underlying pathobiology of asthma, and specific manifestations of asthma such as allergen or exercise-induced responses, while the direct challenge test with methacholine is a sensitive test that serves as a more general indicator of AHR that may not be specific for asthma. The prototypical clinical manifestation of indirect AHR is the development of bronchoconstriction in response to exercise challenge, a clinical disorder that is called exercise-induced bronchoconstriction (EIB) [5–7]. In this regard, there is substantial discordance between the severity of EIB and measures of direct AHR such as methacholine challenge. It is common that individuals with asthma do not have EIB but do have AHR to methacholine challenge, and in some cases there is enough discordance that subjects with a negative methacholine challenge have EIB.

Some of the indirect challenge tests have the potential to trigger severe bronchoconstriction, because the challenge test is conducted with a single strong stimulus for bronchoconstriction, in contrast to the methacholine challenge that is conducted in a graded manner from a low initial starting dose. In this technical standard, we review the pathophysiological basis for indirect challenge testing, and the methods to conduct several of the most common tests. The challenge tests that use a single strong hyperpnoea stimulus including exercise challenge and eucapnic voluntary hyperpnoea (EVH) are discussed initially, followed by the methodology for the graded challenge tests including hypertonic saline, mannitol and adenosine challenge. Specific allergen challenge, which is also a form of indirect AHR and is used predominantly as a research tool in specialised centres, is discussed briefly; a recent report that was reviewed by this task force provides a detailed description of specific allergen challenge [8].

Methods

This document is the second of two technical standards that were developed by a task force on bronchial challenge testing initially organised by the European Respiratory Society (ERS) and American Thoracic

Society (ATS) as outlined in the first technical standard [1]. Due to the extended timeline of the project, this document was completed under ERS-only sponsorship with the participation of the full international panel. The task force initially met and performed a structured literature review on bronchoprovocation testing from 1990 onward. The initial literature review was conducted through PubMed and Embase by University of Washington librarians. The specific search terms were detailed in appendix A of the first report [1], and included terms related to both direct and indirect challenge tests. The initial literature search identified 2235 potentially relevant citations that were reviewed by the chair of the task force, who selected papers that reported methodology, compared methods or reported the outcomes of testing in larger populations, eliminated references that were not relevant and grouped the references by topic. After the initial meeting of the task force, working groups were established who reviewed the literature related to each type of indirect challenge test, specifically those related to exercise, EVH and cold air challenge testing, osmotic challenge testing including hypertonic saline and mannitol challenge tests and adenosine challenge. Task force members initially reviewed the citations for potential inclusion in the document, and then subsequently updated that search in 2017 prior to the completion of each section.

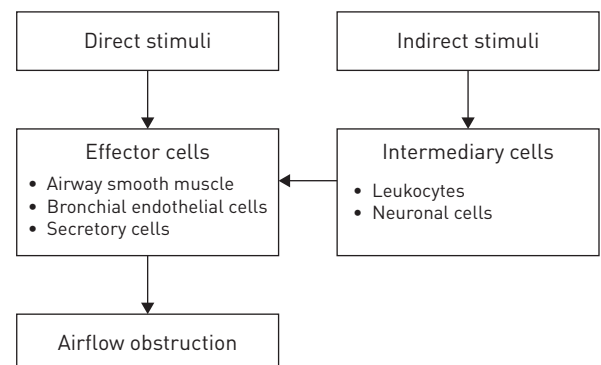
370 documents were found to be relevant to the indirect challenge testing technical standard. This included 42 documents related to adenosine challenge, 32 related to hypertonic saline challenge, 66 related to mannitol challenge, 207 related to exercise challenge, 38 related to EVH, and 23 related to cold air challenge. A task force member with specific expertise in each area reviewed the documents for potential inclusion in the final technical standard and a subset of these references were included in the final document.

Pathophysiological basis and rationale

Ventilation with large amounts of air during exercise and other related stimuli will trigger EIB in ~30–60% of patients with asthma [9–13]. Cross-sectional screening studies have found rates of EIB between 10% and 18% in the general population, suggesting that EIB may be more common than previously recognised in some populations and that certain individuals who have not been diagnosed with asthma may have this disorder [14, 15]. Atopy is a recognised risk factor for EIB, suggesting that subjects with allergic rhinitis or atopic dermatitis with exercise-related symptoms may have EIB [16–19]. There is evidence that EIB is a manifestation of airway inflammation since patients with EIB, relative to asthmatics without this aspect of asthma, have higher levels of cysteinyl leukotrienes (CysLTs) [20, 21], and 8-isoprostanes [22] in their airways; furthermore, the level of exhaled nitric oxide is also correlated with the severity of EIB [23–28]. A recent quantitative immunopathology study revealed that the density of intra-epithelial mast cells is increased in subjects with asthma who have EIB as compared with subjects with asthma who do not have EIB and nonasthmatic controls [29]. In addition, there is a relationship between the percentage of sputum eosinophils and the severity of EIB [30]. Some studies indicate that EIB and other features of asthma occur frequently in high-level athletes, such as cross-country skiers without a prior history of asthma, after exposure to training environments in which they inspire large volumes of cold dry air that may result in injury to the airways [31–36]. Identification of this disorder is particularly important, since asthma triggered on the playing field was the most common cause of nontraumatic death during sports in a community-based survey [37].

Although the susceptibility to EIB varies markedly among subjects, the severity of bronchoconstriction induced on any one occasion is strongly related to the amount of ventilation achieved, up to a maximal plateau [38], as well as the water content and temperature of the inspired air. During periods of high ventilation, a large volume of air is equilibrated to the humidified conditions of the lower airways over a short period of time, leading to the transfer of both water and heat to the inspired air with resulting

FIGURE 1 Conceptual framework for the difference between direct and indirect challenge tests. Indirect challenge tests act indirectly through the activation of leukocytes and/or neuronal cells that lead to the subsequent development of airflow obstruction. Reproduced and modified from [3] with permission from the publisher.



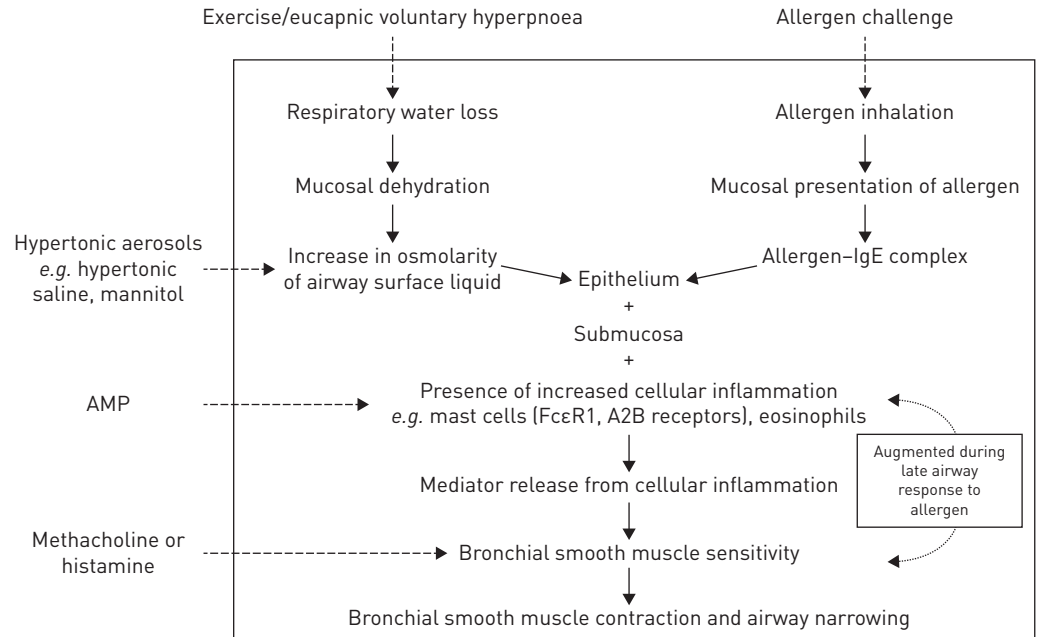


FIGURE 2 Mechanisms of action of indirect challenge tests used in the clinical and research setting. In contrast to the indirect mechanisms of indirect airway hyperresponsiveness, methacholine and histamine are examples of direct challenge tests that cause airway narrowing directly through airway smooth muscle contraction. Reproduced and modified from [4] with permission from the publisher.

osmotic stress as well as cooling of the airways that results from this vaporisation of water [5]. The role of cooling in isolation remains controversial, since very cold temperatures are required to accentuate airway narrowing, such cold air is not required for bronchoconstriction and EIB can occur when the inspired air is above body temperature [5]. Although the precise nature of the initial stimulus remains incompletely understood, there is strong evidence that mediators including CysLTs and prostaglandin D₂ are released into the airways in response to exercise challenge [39–44] and that the mediators that are released are derived from cells residing in the airways, including mast cells and eosinophils [40, 41, 45]. Sensory nerve involvement has been clearly demonstrated following hyperpnoea-induced bronchoconstriction in animal models [46, 47] and studies in humans are supportive of sensory nerve involvement [48]. Despite these inflammatory events triggered by exercise, there is no clear evidence of a cellular influx into the airways or an increase in direct AHR following exercise challenge [49].

Indications

Exercise challenge is the most straightforward method to determine whether exercise-related respiratory symptoms represent EIB rather than dysfunctional breathing, airflow obstruction related to chronic asthma or other cardiovascular pathology. In addition, patients with EIB generally demonstrate AHR to methacholine; however, there is significant discordance between AHR to methacholine and the severity of EIB [29, 50, 51]. Although supportive of the potential diagnosis of EIB, a positive methacholine challenge test does not rule in EIB, and a negative methacholine challenge does not entirely exclude EIB [50, 52, 53], as some athletes with a positive hyperpnoea challenge test have a negative methacholine challenge [51, 54]. In general, AHR to methacholine is a test that is sensitive for asthma but is not very specific, while EIB is less sensitive and more specific for asthma [13, 53, 55–59]. The sensitivity of indirect challenge tests varies with the population that is being studied, as one multicentre trial found that in subjects with relatively normal lung function, mild symptoms and mild AHR, direct and indirect tests had similar sensitivity for a clinical diagnosis of asthma and for detection of EIB [52].

The knowledge that an individual subject has EIB can be used to guide therapy targeted at prevention of symptoms and signs related to this disorder. Previously, testing for EIB or asthma was required by some governing bodies to obtain approval for the use of certain asthma medications during athletic competition. In subjects engaged in demanding or lifesaving work that may require intense exercise (e.g. military, police or firefighting work), a test for EIB may be indicated [60]. In children, identification and treatment of EIB may be particularly relevant because of the important effects of physical activity for physical and psychological development [61]. As a model stimulus of indirect AHR, exercise challenge can serve as a means to study the efficacy and optimal dose of preventative therapy for asthma [62].

Safety considerations and contraindications

Unlike progressive challenge tests such as the methacholine or mannitol challenge tests with an incremental increase in the dose of the stimulus, exercise, EVH and cold air challenge tests build rapidly to a strong stimulus for bronchoconstriction, making it imperative that the laboratory has appropriate equipment available to manage severe bronchoconstriction. A physician should be present or immediately available during the study, and cardiopulmonary resuscitation equipment should be immediately available. During exercise, arterial oxygen saturation should be estimated by pulse oximetry, and blood pressure and ECG should be monitored. The patient should be observed for undue stress (e.g. severe wheezing or shortness of breath, chest pain, lack of coordination) or adverse signs (e.g. ECG abnormalities, falling blood pressure, decrease in oxygen saturation) during the test. Because of the risk of severe bronchoconstriction, it is recommended that the forced expiratory volume in 1 s (FEV₁) before challenge should be $\geq 75\%$ predicted and pulse oximetry saturation should be $>94\%$ [63]. Additional contraindications include significant cardiovascular disease, such as inducible cardiac ischaemia, uncontrolled hypertension, aortic aneurysm or life-threatening arrhythmias (see table 1 in COATES *et al.* [1]). A 12-lead ECG should be obtained in subjects with cardiovascular risk factors, while a three-lead ECG may be acceptable in individuals with a low pretest probability of heart disease and in children. Pregnancy is a contraindication to testing due to the potential risk to the fetus. Safety recommendations are based on physiological considerations and on substantial experience reported in the literature [60, 63].

Patient preparation

The patient should report to the laboratory in comfortable clothes and running or gym shoes, having consumed no more than a light meal. Both short- and long-term medications should be withheld before testing to prevent the possibility of a false negative test [7] (table 1), unless the goal is to assess the efficacy of therapy to prevent EIB [64]. It should be noted that regular use of β_2 -agonists increases the severity of EIB [65, 66]. Some studies have demonstrated a preventative effect of a single high dose of an inhaled corticosteroid prior to EVH challenge [67] or prior to exercise challenge [68], but most studies have found that lower standard doses of inhaled corticosteroids used daily require a longer period of use for optimal efficacy [69]. Hence, maintenance therapy with inhaled corticosteroids can suppress EIB [62, 69]. Vigorous

TABLE 1 Withholding times prior to indirect challenge testing

	Withholding time	Maximum duration of protection [#]
SABA (albuterol, terbutaline)	8 h	<6 h
LABA (salmeterol, eformoterol)	36 h	12 h
LABA in combination with an ICS (salmeterol/fluticasone, formoterol/budesonide)	36 h	NA
Ultra-LABAs (indacaterol, olodaterol, vilanterol)	48 h	NA
ICS (budesonide, fluticasone propionate, beclomethasone)	6 h	NA
Long-acting ICS (fluticasone furoate, ciclesonide)	24 h	NA
Leukotriene receptor antagonists (montelukast, zafirlukast)	4 days	24 h
Leukotriene synthesis inhibitors (zileuton/slow-release zileuton)	12 h/16 h	4 h
Antihistamines (loratadine, cetirzine, fexofenadine)	72 h	<2 h
Short-acting muscarinic acetylcholine antagonist (ipratropium bromide)	12 h	<0.5 h
Long-acting muscarinic acetylcholine antagonist (tiotropium bromide, aclidinium bromide, glycopyrronium)	72 h	NA
Cromones (sodium cromoglycate, nedocromil sodium)	4 h	2 h
Xanthines (theophylline)	24 h	NA
Caffeine	24 h	NA
Vigorous exercise	4 h	<4 h

Examples of specific medications within the class are provided in parenthesis. The withholding times recommended for short- and long-acting β_2 -agonists differs from the recommendation in the original reference, reflecting the recommendations of this task force and the uncertainty in the precise duration of the inhibitory effect of these medications on indirect airway hyperresponsiveness. SABA: short-acting β_2 -agonist; LABA: long-acting β_2 -agonist; ICS: inhaled corticosteroid; NA: not available. [#]: refers to the potential effects of a single dose and may not apply to chronic dosing. Reproduced and modified from [7] with permission from the publisher.

exercise should be avoided for ≥ 4 h before testing, because a “warm up” exercise may cause a period where the subject is refractory to additional exercise challenge [60, 70, 71]. The time of day should be consistent in repeated studies, as the severity of EIB is greater in the afternoon compared to the morning, indicating an influence of diurnal variability [72]. In addition, dietary factors including a low salt diet [41] and supplemental omega-3 fatty acids [43, 44], as well as antioxidants [73], may influence EIB severity.

Prior to any of the challenge tests, it is critical to obtain reliable baseline spirometry since the response to exercise, EVH or cold air challenge is based on the change in FEV₁ from baseline. Accordingly, baseline spirometry should be obtained following ATS/ERS guidelines immediately prior to beginning of the challenge test [74]. It is ideal to obtain two separate baseline spirometry tests separated by 10–20 min before the challenge test to confirm the stability of spirometry.

Exercise challenge testing

Exercise testing can be accomplished using a motorised treadmill or a cycle ergometer. The rapid increase in ventilation during treadmill running makes it the preferable test; however, a cycle ergometer can be used effectively, providing that the work rate is increased rapidly to reach the target ventilation or heart rate. Although peak oxygen uptake averages $\sim 10\%$ less on a cycle ergometer than on treadmill, the lung function response is comparable as long as similar exercise intensity and ventilation are achieved [60, 75]. An outdoor free-run asthma screening test has been used in the community setting to screen for EIB [76–80], although environmental conditions are hard to standardise and safety measures are more difficult to provide in this setting. Sports-specific tests have also been applied in competitive athletes [81, 82].

Regardless of the mode of exercise test, the protocol should be designed to reach the target heart rate or minute ventilation over a short period of time, in the order of 2–3 min (table 2). The rapid rise in work rate is needed because a warm-up period or prolonged lower-level exercise may decrease the severity of EIB [70, 71, 83]. For this reason, the incremental work rate profile used in standard cardiopulmonary exercise testing may be less sensitive for the detection of EIB [60]. Heart rate and rhythm should be monitored with a continuous ECG, and an initial rise in blood pressure with exercise should be confirmed.

Following the rapid increase in work, the target level should be maintained for 6 min [60, 63]. If the equipment is available, it is preferable to achieve a ventilation target rather than a heart rate target to monitor the intensity of the challenge. Ventilation should reach 60% of the predicted maximum voluntary ventilation (MVV, estimated as FEV₁ \times 40) [63, 84]. Monitoring ventilation requires the patient to breathe through a mouthpiece or mask, which adds some burden to conducting the test. An acceptable alternative is a target heart rate of $>85\%$ of the predicted maximum (calculated as 220 – age in years) [84, 85]; however, the approach based on heart rate may not always achieve the target ventilation [86], and higher heart rate targets can be associated with more bronchoconstriction [83]. Measurement of pulmonary gas exchange during exercise makes it possible to quantify the intensity of exercise as a fraction of predicted peak oxygen uptake, confirming the adequacy of exercise challenge. However, adding pulmonary gas exchange measurements is difficult when using a dry air setup and is usually not necessary.

The inspired air should be relatively dry and $<25^\circ\text{C}$. This can be accomplished by conducting the study in an air-conditioned room (with ambient temperature at 20–25°C) with low relative humidity ($\leq 50\%$). The temperature and relative humidity should be recorded. An ideal system delivers dry air through a mouthpiece and a two-way valve from a talc-free reservoir filled with medical-grade compressed air [60]. The use of compressed air is preferred because it is completely dry and will cause greater water loss from

TABLE 2 Characteristics of hyperpnoea challenge tests

Vigorous exercise, usually on a treadmill

Rapid increase in ventilation over the first 2–3 min to reach the target

Maintain target ventilation for ≥ 4 min, preferably 6 min

Target ventilation is 60% of maximum (MVV or FEV₁ \times 40)

Heart rate of $>85\%$ of maximum can serve as a surrogate for ventilation target

Inspired air should be dry, and ambient temperature $<25^\circ\text{C}$

Nose clips should be used during the exercise challenge

Serial assessments of spirometry for 30 min after exercise

Two spirometry manoeuvres are acceptable at each time point

Use the best technically valid FEV₁ at each time point

Frequent deep inspiration following exercise challenge may affect result

MW: maximum voluntary ventilation; FEV₁: forced expiratory volume in 1 s.

the airways, thus generally increasing the sensitivity of the test [38]. This may be particularly important for the detection of mild EIB, and during tests that are of shorter than the ideal 6-min plateau phase. During exercise, the patient should wear a nose clip, as nasal breathing decreases water loss from the airways [60].

Treadmill protocol

On the treadmill, speed and grade are progressively advanced during the first 2–3 min of exercise until the target level is obtained. The degree of physical fitness and body weight will strongly influence the grade and speed necessary to obtain the desired ventilation or heart rate. A reasonable procedure is to quickly advance to a rapid, but comfortable speed at a treadmill incline of 5.5% (3°) then, raise the slope until the desired heart rate or ventilation is obtained, up to an incline of 10%. Nomograms have been proposed to predict speed and grade that will elicit the desired heart rate, but they have not been validated extensively [60]. Another approach is to use a nomogram relating oxygen consumption per kilogram to speed and slope of the treadmill; however, the relationship between oxygen consumption and the percentage of maximum ventilation is variable [60]. The test ends when the patient has exercised at the target ventilation or heart rate for 6 min. The treadmill challenge protocol has a high degree of repeatability [87–89].

Bicycle ergometer protocol

For cycle ergometer exercise, work rate is rapidly increased using the electromagnetic braking system to achieve the target ventilation. Direct measurement of ventilation is easier with the stable position on an ergometer and is the preferred target. The target heart rate or ventilation should be reached within 2–3 min. A valid test requires the target exercise intensity to be sustained for 6 min, although sustained exercise of ≥ 4 min may be acceptable if the subject becomes fatigued. Although the repeatability of the bicycle protocol has not received extensive study, the repeatability in a limited number of individuals was excellent [60, 90].

Eucapnic voluntary hyperpnoea

EVH is an alternative to exercise challenge that was standardised in the early 1980s [91, 92] and does not require exercise. The study utilises medical dry air from a reservoir with an admixture of 4.9% carbon dioxide, which enables the study subject to breathe at high ventilation without the adverse consequences of hypocapnia. The subject is instructed to perform voluntary hyperpnoea for 6 min aiming at a target ventilation of 85% of MVV and with a minimum ventilation threshold of 60% of MVV [93, 94].

Like other indirect challenge tests, EVH has a lower sensitivity for asthma compared to direct tests such as methacholine challenge, but is fairly specific for asthma [6]. Like exercise challenge, the volume and water content of the inspired air are important determinants of the severity of bronchoconstriction following EVH [94, 95], and the use of certain asthma medications before the challenge can alter the sensitivity and specificity of the test (table 1) [67]. When standardised properly, the EVH test has a high degree of repeatability [95]. This method has been used to identify the origin of exercise-related symptoms in subjects with respiratory disorders [96], in elite athletes [97], in school-aged athletes [98] and in people with a health club membership [99]. In addition, EVH has been used to assess the efficacy of inhaled corticosteroids in infants and young children [100]. When compared to exercise challenge, a consistent finding is a higher sensitivity of EVH than exercise challenge for the detection of bronchoconstriction [81, 97, 101, 102].

Cold air challenge

Cold air has a low water-carrying capacity, resulting in greater heat and water transfer necessary to condition the inspired air at any minute ventilation. Since this greater heat and water flux is thought to be instrumental in the pathogenesis of EIB, performing exercise or EVH with cold air inhalation might be expected to increase the stimulus for bronchoconstriction. Cold air generators that produce dry air at below-freezing temperatures are commercially available and are in use in some laboratories; however, the additive effects of cold air depend on the specific protocol that is used. In adults, some studies demonstrate an enhancing effect of cold compared to ambient air [103], while others found no additive effect of cold air [104, 105]. An increase in minute ventilation elicited by the use of cold air explains some of the effects of cold air [106]. In children, cold air inhalation enhances the response to EHV in those with asthma [55, 107]. The addition of cold air appears to shorten the stimulus time necessary during challenge testing from 6 to 4 min [84]. In patients with symptoms specifically associated with exercise in the cold, exercise challenge while breathing cold dry air may be useful to enhance the sensitivity and specificity of the response [82, 97]. Part of the response to exercising in cold temperatures may be due to exposure of the face and body to cold temperatures, and not just the airways [108, 109]. Using cold air as part of an

exercise or EVH challenge elicits responses that are discordant from direct challenge tests such as methacholine or histamine [110, 111].

An exercise or EVH challenge using cold air is conducted as previously described, but with a source of frigid air for inspiration. Typically, subjects breathe through a heat exchanger or similar device that generates cold, dry air or exercise under ambient conditions of cold air, such as might be found outdoors during the winter or inside an ice skating arena [97]. When using a cold air generator, the device is either held by the patient or is supported in such a way as to deliver the air immediately before inspiration. The target range for inspired air temperature is -10 – -20°C , and should be recorded by the technologist during the challenge.

Selection of the hyperpnoea challenge protocol

Although all the methods are supported in the literature as valid challenge tests for EIB, we recommend the use of the treadmill protocol monitoring heart rate as a surrogate for ventilation because of the wide availability of this method and general familiarity to both the physician and the patient with running on a treadmill. The other methods are also acceptable but require the use of specialised equipment (*e.g.* bicycle ergometer in bike challenge, feedback mechanism to maintain target ventilation in EVH, cold air generator) or protocols.

Post-challenge spirometry protocol

Serial measurements of lung function by FEV₁ over the first 30 min after exercise or EVH are used to determine whether the test is positive and quantify the severity of bronchoconstriction. Many laboratories conduct the first spirometry measurements immediately after challenge, and then 3, 6, 10, 15 and 30 min after challenge. An acceptable alternative is to start the assessment at 5 min after challenge; however, the earlier time intervals may be useful to detect the onset of severe bronchoconstriction if present at the cessation of the challenge test [60, 84]. Note that when performing spirometry measurements immediately after stopping exercise, the result might be artificially low due to reduced effort during spirometry in the setting of post-exercise hyperpnoea and dyspnoea, particularly in children. There may be some slight differences in the results when different numbers of spirometry manoeuvres are used, since deep inspiration during spirometry testing may inhibit bronchoconstriction [112]. It should be noted that the bronchoprotective effects of a deep inspiration is reduced in asthma and has been related to inflammation, suggesting that deep inspiration could add to the discriminant value of the tests [113–115]; however, we feel that the number of deep inspirations for the spirometry manoeuvre should be kept as constant as possible to minimise the variability of the test. At least two acceptable tests within 0.15 L should be obtained at each testing interval, unless FEV₁ or forced vital capacity are <1 L, in which case two acceptable tests should be within 0.10 L. The best FEV₁ at each interval is reported. Because of the challenge in carrying out repeated spirometry efforts following exercise, it is adequate to accept a duration of expiration of 2–3 s for the forced manoeuvre to measure FEV₁; however, it is important to vigorously coach the patient to inhale fully even in the presence of chest tightness.

In most cases, the nadir in FEV₁ occurs within 5–10 min of cessation of exercise, but can occasionally occur as late as 30 min post-exercise [116]. We recommend serial measurements for the entire 30 min since it is important to determine the nadir in FEV₁ to fully assess the severity of EIB.

Positive test threshold and interpretation of hyperpnoea challenge tests

The presence of EIB is defined by plotting FEV₁ as a percentage decline from the pre-exercise baseline FEV₁ at each post-exercise interval. A decrease of $\geq 10\%$ from the baseline FEV₁ is accepted as an abnormal response relative to population normal values [63, 116–118], but the specificity is higher with a criterion of 15% from baseline. The basis of these recommendations are studies in normal children that demonstrate an upper 95% confidence limit of the FEV₁ fall as 8.2% [118] and 10% [119]. A method to quantify the overall severity of EIB is to measure the area under the curve for post-exercise time multiplied by the percentage fall in FEV₁ at each time point over the 30 min after exercise [85]; this quantitative assessment is used predominantly in the research setting as a specific threshold for a positive test has not been established. The threshold for a positive response based on the maximum fall in FEV₁ also depends on the indication for the test, such that a more sensitive test (*i.e.* 10%) might be useful to understand the origin of symptoms in athletes, while a more specific test (*i.e.* 15–20%) may be needed for research studies [7].

For EVH, the threshold for a positive test is usually set at a decrease in FEV₁ of $\geq 10\%$ below baseline based on the characteristics of the EVH test in normal subjects [96]. Exercise or EVH challenge tests that include the addition of cold air should be interpreted in the same manner as the test conducted without cold air.

A β_2 -agonist bronchodilator may be administered at any time to reverse bronchoconstriction if the patient experiences symptoms that are too severe, if there is severe bronchoconstriction that could progress further or if FEV₁ has not recovered to within 10% of baseline when the patient is ready to leave the laboratory. The clinician should be alert to other possible causes of dyspnoea including cardiovascular disease and upper airway abnormalities including fixed upper airway obstruction (*i.e.* subglottic stenosis) and inducible laryngeal obstruction including paradoxical vocal cord dysfunction and paradoxical arytenoid motion [120]. Upper airway abnormalities may be apparent if full flow–volume loops with inspiratory flow are obtained during spirometry; however, other techniques, such as direct laryngoscopy immediately after or during exercise may be needed to clarify the diagnosis [120].

Incremental indirect airway challenge tests

In addition to the hyperpnoea challenge tests that use a single strong stimulus for bronchoconstriction, several incremental challenge tests have been developed to measure indirect AHR. As water transfer during a period of hyperpnoea results in osmotic stress to the lower airways, the inhalation of aerosols with different osmotic properties than the airway surface liquid has been developed to model this stimulus in an incremental manner [121]. The response to either hypertonic or hypotonic aerosols is associated with the response to exercise [6, 122, 123] and EHV [124] challenge tests, and the response to hypertonic aerosols is also associated with mast cell infiltration of the airways and sputum eosinophils [125, 126]. Challenge with hypertonic saline causes mediator release following challenge that is similar to that described for exercise challenge [127] and can be inhibited by specific mediator antagonists [128]. In model systems, hypertonic saline leads to the release of neuropeptides that are involved in the development of bronchoconstriction [129]. Similarly, the use of dry powdered mannitol has been developed as an incremental challenge test that challenges the lower airways with increasing levels of osmotic stress [130]. A positive response to mannitol challenge is generally associated with a positive test for EIB [131, 132], but mannitol challenge is neither entirely sensitive or specific for the detection of EIB [52, 133]. Additionally, a recent study demonstrated that AHR to mannitol is associated with mast cell and eosinophil infiltration of the airways [134], consistent with prior evidence of the release of mast cell mediators following mannitol challenge [39], and a reduction in AHR to mannitol with a mast cell stabiliser [135]. Another incremental challenge test to define indirect AHR is the dose response to inhaled adenosine. Although not extensively studied, the response to adenosine challenge is generally correlated with the response to exercise challenge [136]. Furthermore, the response to adenosine challenge is associated with markers of airway inflammation, including the exhaled nitric oxide fraction [137, 138]. The mechanism of adenosine-induced AHR may be through the priming of mast cells [139], adding further evidence of a strong relationship between mast cell infiltration of the airways and indirect AHR.

Hypertonic saline challenge

Although the inhalation of either hypertonic or hypotonic aerosols will induce bronchoconstriction in susceptible individuals, the use of hypotonic aerosols is uncommon, due in part to a fatal episode of asthma triggered during distilled water challenge [140]. Hypertonic saline challenge is an incremental challenge in which 4.5% sterile saline is delivered by an ultrasonic nebuliser for increasing periods of time [124, 141]. The test should be conducted with an ultrasonic nebuliser with a flow of ≥ 1.2 mL·min⁻¹ and the capacity to hold ≥ 100 mL hypertonic saline solution. The commonly used ultrasonic nebulisers provide a flow of 1.5–3.0 mL·min⁻¹ [142, 143]. Because the “dose” or volume of hypertonic saline required to induce a 15% fall in FEV₁ is the primary outcome measurement, the precise output from the nebuliser should be established gravimetrically by the laboratory conducting the test. Ideally, the hypertonic saline should be delivered through a two-way nonrebreathing valve, and the inner walls of the tubing should be smooth and of constant length from test to test, as the deposition in the tubing has the potential to alter the characteristics of the test. A saliva collection reservoir should be used, as the test tends to cause ongoing salivation. During the test, the sterile 4.5% hypertonic saline is given for increasing durations of time until the longest duration is administered, or there is a >15% fall in FEV₁ from the pretest baseline. If FEV₁ falls between 10% and 15% compared with prechallenge FEV₁, the same dose step is repeated. Spirometry is used to assess the FEV₁ 30 and 90 s after the end of each round of inhaled hypertonic saline. Hypertonic saline is initially administered for 30 s, then 1, 2, 4 and 8 min in subsequent cycles. When the test is positive, or symptoms are present, a short-acting inhaled β_2 -agonist should be administered, and the patient monitored until fully recovered to within 10% of baseline FEV₁.

The contraindications for the test are generally the same as for other indirect challenge tests, with a baseline FEV₁ >70% pred considered safe for this type of test. The test is considered positive if there is a >15% fall in FEV₁ from the prechallenge FEV₁ during the test [123, 124]. The provocative dose causing a 20% fall in FEV₁ (PD₁₅) of hypertonic saline is established by linear interpolation of a plot of the cumulative dose (mL or g) of hypertonic saline administered during the test and the FEV₁ at the end of

each nebulisation cycle. The severity of AHR to hypertonic saline can be classified as mild ($PD_{15} >6$ mL), moderate (PD_{15} 2.1–6.0 mL) and severe ($PD_{15} <2.0$ mL), although precise cut-offs are uncertain, as large population studies have not been conducted [144–146]. Another outcome measure described for this test is the dose response slope (DRS), which is the percentage fall in FEV_1 divided by the amount of hypertonic saline inhaled. In a large study of children, the DRS had greater separation for current wheezing and for asthma than the PD_{15} hypertonic saline [144].

An advantage of hypertonic saline as a test of indirect AHR is the ability to conduct an induced sputum test at the same time, to characterise features of airway inflammation [126, 147]. Although hypertonic saline challenge has not been used extensively in population based studies, there is a strong scientific basis that the response to hypertonic saline is similar to other indirect tests such as exercise challenge [123] and EVH [148]. Population studies, predominantly in children, support the ability of hypertonic saline challenge to discriminate between subjects with and without asthma and related features of asthma [149–151]. Further support for the connection between AHR to hypertonic saline and airway inflammation are studies that reveal that anti-inflammatory therapies including inhaled steroids [126, 145, 147, 152] and cromones [153] alter the severity of AHR to hypertonic saline.

Mannitol challenge testing

The mannitol challenge test has undergone considerable evaluation and has received regulatory approval in the United States and Europe. The test is conducted using a capsule-based dry powdered inhaler device that is used to deliver increasing doses of mannitol to the lower airways. The dry powder is prepared by the manufacturer by spray-drying mannitol into particles that are of respirable size. Inhalation of dry powdered mannitol rapidly increases the osmolarity of the airway surface liquid [130, 131]. After baseline spirometry is obtained, a capsule without any mannitol is inhaled through the delivery system. The capsule is placed in the inhaler and pierced by simultaneously depressing the two buttons on each side of the dry powder inhaler. The individual is instructed to exhale fully away from the inhaler, tilt the head back slightly and place the inhaler in their mouth tilted upward at a 45° upright angle with lips sealed around the mouthpiece, and then take a slow deep inhalation. The subject should be instructed to hold their breath for 5 s after inhalation. Nose clips may be used if desired. During inhalation, a “rattling” sound should be heard as the capsule spins within the inhaler. When 5 s has passed, the patient is instructed to exhale through the mouth (away from the inhaler), to remove the nose clip, and to breathe normally. 1 min after the start of inhalation, two acceptable spirometry manoeuvres should be obtained, with the highest FEV_1 retained as the baseline FEV_1 . Following spirometry, increasing doses of mannitol (5, 10, 20, 40, 2×40, 4×40, 4×40 and 4×40 mg) are administered using the same cycle of the inhaled dose from the dry powder inhaler, followed by two acceptable spirometry manoeuvres conducted 1 min after administration of the dose. When multiple capsules are required for the dose, the timer is started at the start of inhalation of the final dose. Because there is a cumulative dose effect, the next dose should be administered as soon as possible following spirometry, and delays between doses should be avoided. The test is stopped when there is a 15% decrease from baseline FEV_1 , a 10% decrease in FEV_1 between two consecutive doses or the cumulative dose reaches the total of 635 mg.

The contraindications for mannitol challenge testing are like those for other indirect challenge tests, with a few additional considerations. First, known hypersensitivity to mannitol or to the gelatine used to make the capsules is an absolute contraindication. Second, moderate airflow limitation ($FEV_1 <70\%$ pred or <1.5 L) is considered a contraindication to testing. As cough is frequently induced by inhaled mannitol, the test should be avoided in patients with medical conditions that would be exacerbated by frequent coughing. Finally, as mannitol is a pregnancy category C drug and is a drug with potential excretion in breastmilk, administration of mannitol challenge should be avoided in pregnant women and in nursing mothers. Following mannitol challenge with a positive test or significant respiratory symptoms (e.g. wheezing, dyspnoea, cough), a short-acting inhaled β_2 -agonist should be administered, and the patient monitored until fully recovered to within 10% of baseline FEV_1 . In the case of a negative result, if the patient has significant respiratory symptoms, a short-acting inhaled β_2 -agonist may be considered.

A mannitol challenge test is considered positive if there is a 15% decrease from baseline FEV_1 in response to the cumulative total dose, or a 10% decrease in FEV_1 between two consecutive mannitol doses. The test is considered negative when a cumulative dose of 635 mg of mannitol has been administered and the FEV_1 has not fallen by $\geq 15\%$ from baseline. Sensitivity to mannitol is expressed as the PD_{15} derived from the dose-response curve (PD_{15} mannitol). The PD_{15} mannitol can be used to classify the severity of indirect AHR as mild (>155 mg), moderate (>35 and ≤ 155 mg) or severe (≤ 35 mg) (figure 3) [84, 154, 155]. Reactivity to mannitol can also be expressed as the response dose ratio (RDR), which is the percentage decrease in FEV_1 at the end of challenge divided by the cumulative dose of mannitol administered to induce that decrease in FEV_1 [130, 143]. The PD_{15} is generally used in clinical practice to “rule in” or

establish a diagnosis of asthma, and for providing an estimate of the severity of AHR. The RDR makes it possible to include subjects who do not reach the cut-off in the analysis, providing a continuous measurement of indirect AHR without censored data, which may make it particularly useful for epidemiological studies as has been shown for similar methods of assessment of direct AHR [156, 157]. Additionally, the mannitol challenge test has utility as a screening test for asthma [158, 159], to monitor the effectiveness of long-term controller therapy for asthma [160–163], and to determine the optimal dose of inhaled corticosteroid therapy [164–166]. In cross-sectional studies in a clinical setting, the sensitivity of the mannitol bronchial challenge test to identify asthma ranges from ~40% to 59%, while the specificity ranges from ~78% to nearly 100% [52, 159, 167].

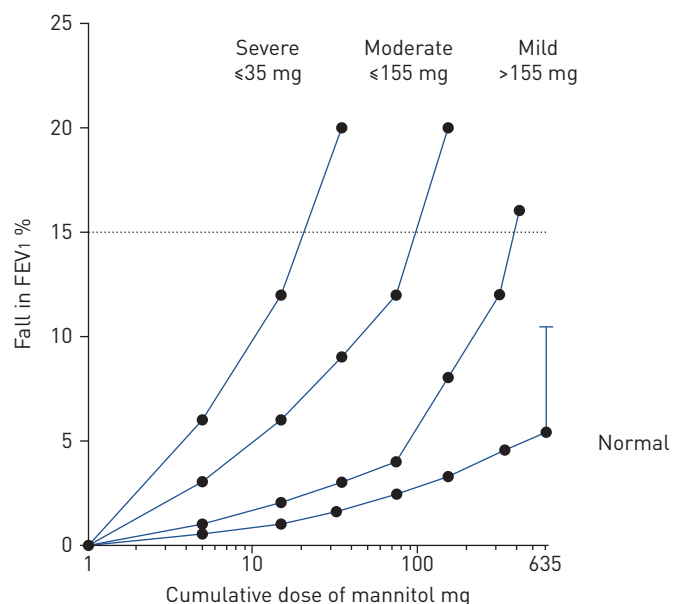
Adenosine challenge testing

The adenosine bronchial challenge test is used primarily in the research setting, because specific thresholds for the interpretation of the test have not been clearly established. The adenosine challenge test is usually performed with an inhaled solution of AMP because AMP is more soluble in sterile normal saline than adenosine, and is rapidly metabolised to adenosine in the lungs [168]. The sodium salt of AMP is available as a dry crystalline powder from chemical suppliers; however, pharmaceutical grade AMP is not currently available for this test, and should only be used with appropriate regulatory approval. The use of a 2-min tidal breathing as well as a five-breath dosimeter protocol with doubling concentrations of AMP ranging from 3.125 to 400 mg·mL⁻¹ were described in the 2003 ERS task force on indirect airway challenges [2]. Because of the higher output of modern nebulisers, a lower starting concentration of AMP should be considered [169]. The test is stopped once a 20% fall in FEV₁ occurs and is generally reported as the PC₂₀ AMP. Safety considerations are like those for other indirect challenge tests.

As in the case of methacholine challenge [1], there are several important considerations that affect the standardisation and interpretation of the adenosine challenge test. First, the well-described bronchoprotective effect of deep inspiration may alter the characteristics of the tests when the dosimeter method is used. Second, modern nebulisers have the potential to deliver much higher doses of the drug to the lower airways, such that the dose rather than the concentration should be considered to better standardise the test based on the delivery characteristics of the nebuliser. Consistent with these considerations, one study demonstrated a lower PC₂₀ AMP using the tidal breathing method over the dosimeter method [170]. Another consideration is that the relatively high concentrations of AMP in this dosing scheme have been shown to alter nebuliser delivery during the tidal breathing cycle [171]. A novel delivery system was recently described using two different dry powdered formulations of adenosine; preliminary testing in a modest number of subjects with and without asthma has been reported [172, 173].

Additional studies are needed to fully standardise the delivery of adenosine for the adenosine bronchial challenge test, and to establish well defined thresholds and predictive values for a positive and negative test. The initial studies of AMP challenge indicate that it more closely reflects active airway inflammation in patients with asthma than direct challenge tests [137, 174, 175], and may have a stronger relationship

FIGURE 3 Classification of the severity of airway hyperresponsiveness according to the response to dry powdered mannitol challenge. The cumulative dose of mannitol required to provoke a 15% reduction in forced expiratory volume in 1 s (FEV₁) from baseline is calculated based on the final two cumulative doses of mannitol and the reduction in FEV₁ at each dose. The mean maximum response (+1 sd) in FEV₁ after a cumulative dose of 635 mg is shown in the error bar for normal subjects. Reproduced and modified from reference [84] with permission from the publisher, including data from the normal control group in reference [155].



with the bronchodilator response [176]. Like other indirect challenge tests, the adenosine challenge test may be particularly useful to ascertain the effects of anti-inflammatory therapies such as inhaled steroids in asthma [177–181].

Inhaled allergen challenge

Inhaled allergen challenge is a specific indirect stimulus in allergen-sensitised asthmatics and is used primarily as a model of antigen-specific T-helper type-2 cell-driven asthma, allowing investigation of the links between allergen-induced inflammatory events and subsequent changes in airway physiology [182, 183]. More recently, allergen challenge has been used to define aspects of the innate immune response to allergen [184, 185]. Although sometimes used clinically to demonstrate the relationship between relevant stimuli in symptomatic subjects, the allergen challenge is primarily a research tool, often used to assess the efficacy of novel potential asthma-controlling medications [8, 186]. For the sake of safety and reproducibility, allergen challenges should only be performed in specialised research centres with ample expertise and experience with the methodology of allergen challenge and asthma management. To increase safety, incremental inhalations of an allergen based on the gradual decline in FEV₁ during the early airway response to allergen is recommended. Irrespective of the inhalation method, allergen should only be administered in asthmatics where prechallenge clinical stability is confirmed by strict spirometry and AHR criteria [8].

Summary

The assessment of AHR is exceedingly valuable as a specific diagnostic test for asthma, to understand the basis for asthma symptoms and to recognise the underlying asthma phenotype and thereby to guide and monitor therapy. Direct challenge tests including methacholine challenge have the advantage of a high level of sensitivity, and are therefore most useful to exclude the diagnosis of asthma in the presence of asthma-like symptoms [1]; however, direct AHR is not specific for asthma and can be present in other airway disorders. Depending upon the patient population being examined, indirect tests can have similar sensitivity to methacholine challenge for the detection of asthma [52]. The major strength of indirect challenge tests is that they are more specific for asthma, and can reveal important aspects of the pathophysiology of asthma, and the response to a specific stimulus such as exercise or allergen challenge.

The most commonly used indirect challenge tests, such as exercise challenge and EHV, have a single strong stimulus for bronchoconstriction and need to be used with caution in individuals with symptoms suggestive of poorly controlled asthma or with reduced lung function. Incremental indirect challenge tests, such as hypertonic saline and mannitol challenge, have been developed that can be used to assess indirect AHR across a larger spectrum of individuals with asthma. The assessment of indirect AHR can be useful in several different ways, including diagnostic testing for asthma, epidemiological studies of asthma, and the selection of asthma therapy. As in the case of direct AHR testing, it is crucial that the results of the test are interpreted in the appropriate clinical context and in relation to the features of asthma.

Conflict of interest: T.S. Hallstrand has nothing to disclose. J.D. Leuppi has nothing to disclose. G. Joos reports grants and personal fees for advisory board work and lecturing from AstraZeneca and Novartis, grants from Boehringer Ingelheim and Chiesi, grants and personal fees for advisory board work from GlaxoSmithKline, and personal fees for lecturing from Teva, outside the submitted work. G.L. Hall has nothing to disclose. K.H. Carlsen has nothing to disclose. D.A. Kaminsky has received honoraria for teaching at an annual Cardiorespiratory Diagnostics Course from MGC Diagnostics, Inc., outside the submitted work. A.L. Coates has nothing to disclose. D.W. Cockcroft is a member of a medical advisory board for Methapharm, as has received products for research purposes from Aerogen and Pharmaxis, outside the submitted work. B.H. Culver has nothing to disclose. Z. Diamant reports personal fees from Aerocrine, ALK, Aquilon, AstraZeneca, Boehringer Ingelheim, Gilead, HAL Allergy, MSD and Sanofi Genzyme-Regeneron, outside the submitted work. G.M. Gauvreau has nothing to disclose. I. Horvath reports personal fees and non-financial support from AstraZeneca, Berlin-Chemie, Chiesi, Boehringer Ingelheim, Novartis, CSL Behring and Roche, personal fees from Sandoz, GSK, Sager Pharma, Affidea, Orion Pharma and Teva, and non-financial support from MSD, outside the submitted work. F.H.C. de Jongh has nothing to disclose. B.L. Laube has nothing to disclose. P.J. Sterk has nothing to disclose. J. Wanger has nothing to disclose.

References

- 1 Coates AL, Wanger J, Cockcroft DW, *et al.* ERS technical standard on bronchial challenge testing: general considerations and performance of methacholine challenge tests. *Eur Respir J* 2017; 49: 1601526.
- 2 Joos GF, O'Connor B, Anderson SD, *et al.* Indirect airway challenges. *Eur Respir J* 2003; 21: 1050–1068.
- 3 Van Schoor J, Joos GF, Pauwels RA. Indirect bronchial hyperresponsiveness in asthma: mechanisms, pharmacology and implications for clinical research. *Eur Respir J* 2000; 16: 514–533.
- 4 O'Byrne PM, Gauvreau GM, Brannan JD. Provoked models of asthma: what have we learnt? *Clin Exp Allergy* 2009; 39: 181–192.

- 5 Hallstrand TS. Approach to the patient with exercise-induced bronchoconstriction. *In: Franklin Adkinson N, Bochner BS, Wesley Burks A, et al., eds. Middleton's Allergy Principles and Practice. Philadelphia, PA, Elsevier, 2013; pp. 938–950.*
- 6 Parsons JP, Hallstrand TS, Mastrorarde JG, *et al.* An official American Thoracic Society clinical practice guideline: exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 2013; 187: 1016–1027.
- 7 Weiler JM, Brannan JD, Randolph CC, *et al.* Exercise-induced bronchoconstriction update-2016. *J Allergy Clin Immunol* 2016; 138: 1292–1295.
- 8 Diamant Z, Gauvreau GM, Cockcroft DW, *et al.* Inhaled allergen bronchoprovocation tests. *J Allergy Clin Immunol* 2013; 132: 1045–1055.
- 9 Cabral AL, Conceição GM, Fonseca-Guedes CH, *et al.* Exercise-induced bronchospasm in children: effects of asthma severity. *Am J Respir Crit Care Med* 1999; 159: 1819–1823.
- 10 Kawabori I, Pierson WE, Conquest LL, *et al.* Incidence of exercise-induced asthma in children. *J Allergy Clin Immunol* 1976; 58: 447–455.
- 11 Lazo-Velázquez JC, Lozada AR, Cruz HM. Evaluation of severity of bronchial asthma through an exercise bronchial challenge. *Pediatr Pulmonol* 2005; 40: 457–463.
- 12 Benarab-Boucherit Y, Mehdioui H, Nedjar F, *et al.* Prevalence rate of exercise-induced bronchoconstriction in Annaba (Algeria) schoolchildren. *J Asthma* 2011; 48: 511–516.
- 13 Park HK, Jung JW, Cho SH, *et al.* What makes a difference in exercise-induced bronchoconstriction: an 8 year retrospective analysis. *PLoS One* 2014; 9: e87155.
- 14 Ernst P, Demissie K, Joseph L, *et al.* Socioeconomic status and indicators of asthma in children. *Am J Respir Crit Care Med* 1995; 152: 570–575.
- 15 Priftanji A, Strachan D, Burr M, *et al.* Asthma and allergy in Albania and the UK. *Lancet* 2001; 358: 1426–1427.
- 16 Addo-Yobo EO, Woodcock A, Allotey A, *et al.* Exercise-induced bronchospasm and atopy in Ghana: two surveys ten years apart. *PLoS Med* 2007; 4: e70.
- 17 Caffarelli C, Bacchini PL, Gruppi L, *et al.* Exercise-induced bronchoconstriction in children with atopic eczema. *Pediatr Allergy Immunol* 2005; 16: 655–661.
- 18 Calvert J, Burney P. Effect of body mass on exercise-induced bronchospasm and atopy in African children. *J Allergy Clin Immunol* 2005; 116: 773–779.
- 19 Calvert J, Burney P. Ascaris, atopy, and exercise-induced bronchoconstriction in rural and urban South African children. *J Allergy Clin Immunol* 2010; 125: 100–105.
- 20 Carraro S, Corradi M, Zanconato S, *et al.* Exhaled breath condensate cysteinyl leukotrienes are increased in children with exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2005; 115: 764–770.
- 21 Hallstrand TS, Moody MW, Aitken ML, *et al.* Airway immunopathology of asthma with exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2005; 116: 586–593.
- 22 Barreto M, Villa MP, Olita C, *et al.* 8-Isoprostane in exhaled breath condensate and exercise-induced bronchoconstriction in asthmatic children and adolescents. *Chest* 2009; 135: 66–73.
- 23 Buchvald F, Hermansen MN, Nielsen KG, *et al.* Exhaled nitric oxide predicts exercise-induced bronchoconstriction in asthmatic school children. *Chest* 2005; 128: 1964–1967.
- 24 ElHalawani SM, Ly NT, Mahon RT, *et al.* Exhaled nitric oxide as a predictor of exercise-induced bronchoconstriction. *Chest* 2003; 124: 639–643.
- 25 Lex C, Dymek S, Heying R, *et al.* Value of surrogate tests to predict exercise-induced bronchoconstriction in atopic childhood asthma. *Pediatr Pulmonol* 2007; 42: 225–230.
- 26 Nishio K, Odajima H, Motomura C, *et al.* Exhaled nitric oxide and exercise-induced bronchospasm assessed by FEV₁, FEF_{25–75%} in childhood asthma. *J Asthma* 2007; 44: 475–478.
- 27 Scollo M, Zanconato S, Ongaro R, *et al.* Exhaled nitric oxide and exercise-induced bronchoconstriction in asthmatic children. *Am J Respir Crit Care Med* 2000; 161: 1047–1050.
- 28 Terada A, Fujisawa T, Togashi K, *et al.* Exhaled nitric oxide decreases during exercise-induced bronchoconstriction in children with asthma. *Am J Respir Crit Care Med* 2001; 164: 1879–1884.
- 29 Lai Y, Altmeier WA, Vandree J, *et al.* Increased density of intraepithelial mast cells in patients with exercise-induced bronchoconstriction regulated through epithelially derived thymic stromal lymphopoietin and IL-33. *J Allergy Clin Immunol* 2014; 133: 1448–1455.
- 30 Duong M, Subbarao P, Adelroth E, *et al.* Sputum eosinophils and the response of exercise-induced bronchoconstriction to corticosteroid in asthma. *Chest* 2008; 133: 404–411.
- 31 Medelli J, Lounana J, Messan F, *et al.* Testing of pulmonary function in a professional cycling team. *J Sports Med Phys Fitness* 2006; 46: 298–306.
- 32 Parsons JP, Kaeding C, Phillips G, *et al.* Prevalence of exercise-induced bronchospasm in a cohort of varsity college athletes. *Med Sci Sports Exerc* 2007; 39: 1487–1492.
- 33 Randolph CC, Dreyfus D, Rundell KW, *et al.* Prevalence of allergy and asthma symptoms in recreational roadrunners. *Med Sci Sports Exerc* 2006; 38: 2053–2057.
- 34 Sallaoui R, Chamari K, Mossa A, *et al.* Exercise-induced bronchoconstriction and atopy in Tunisian athletes. *BMC Pulm Med* 2009; 9: 8.
- 35 Bolger C, Tufvesson E, Anderson SD, *et al.* Effect of inspired air conditions on exercise-induced bronchoconstriction and urinary CC16 levels in athletes. *J Appl Physiol* 2011; 111: 1059–1065.
- 36 Bougault V, Turmel J, St-Laurent J, *et al.* Asthma, airway inflammation and epithelial damage in swimmers and cold-air athletes. *Eur Respir J* 2009; 33: 740–746.
- 37 Becker JM, Rogers J, Rossini G, *et al.* Asthma deaths during sports: report of a 7-year experience. *J Allergy Clin Immunol* 2004; 113: 264–267.
- 38 Anderson SD, Schoeffel RE, Follet R, *et al.* Sensitivity to heat and water loss at rest and during exercise in asthmatic patients. *Eur J Respir Dis* 1982; 63: 459–471.
- 39 Brannan JD, Gulliksson M, Anderson SD, *et al.* Evidence of mast cell activation and leukotriene release after mannitol inhalation. *Eur Respir J* 2003; 22: 491–496.
- 40 Hallstrand TS, Moody MW, Wurfel MM, *et al.* Inflammatory basis of exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 2005; 172: 679–686.

- 41 Mickleborough TD, Lindley MR, Ray S. Dietary salt, airway inflammation, and diffusion capacity in exercise-induced asthma. *Med Sci Sports Exerc* 2005; 37: 904–914.
- 42 Hallstrand TS, Chi EY, Singer AG, *et al.* Secreted phospholipase A2 group X overexpression in asthma and bronchial hyperresponsiveness. *Am J Respir Crit Care Med* 2007; 176: 1072–1078.
- 43 Mickleborough TD, Lindley MR, Ionescu AA, *et al.* Protective effect of fish oil supplementation on exercise-induced bronchoconstriction in asthma. *Chest* 2006; 129: 39–49.
- 44 Mickleborough TD, Murray RL, Ionescu AA, *et al.* Fish oil supplementation reduces severity of exercise-induced bronchoconstriction in elite athletes. *Am J Respir Crit Care Med* 2003; 168: 1181–1189.
- 45 Hallstrand TS, Wurfel MM, Lai Y, *et al.* Transglutaminase 2, a novel regulator of eicosanoid production in asthma revealed by genome-wide expression profiling of distinct asthma phenotypes. *PLoS One* 2010; 5: e8583.
- 46 Freed AN, McCulloch S, Meyers T, *et al.* Neurokinins modulate hyperventilation-induced bronchoconstriction in canine peripheral airways. *Am J Respir Crit Care Med* 2003; 167: 1102–1108.
- 47 Lai YL, Lee SP. Mediators in hyperpnea-induced bronchoconstriction of guinea pigs. *Naunyn Schmiedebergs Arch Pharmacol* 1999; 360: 597–602.
- 48 Hallstrand TS, Debley JS, Farin FM, *et al.* Role of MUC5AC in the pathogenesis of exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2007; 119: 1092–1098.
- 49 Hallstrand TS, Altemeier WA, Aitken ML, *et al.* Role of cells and mediators in exercise-induced bronchoconstriction. *Immunol Allergy Clin North Am* 2013; 33: 313–328.
- 50 Henriksen AH, Tveit KH, Holmen TL, *et al.* A study of the association between exercise-induced wheeze and exercise versus methacholine-induced bronchoconstriction in adolescents. *Pediatr Allergy Immunol* 2002; 13: 203–208.
- 51 Holley AB, Cohee B, Walter RJ, *et al.* Eucapnic voluntary hyperventilation is superior to methacholine challenge testing for detecting airway hyperreactivity in nonathletes. *J Asthma* 2012; 49: 614–619.
- 52 Anderson SD, Charlton B, Weiler JM, *et al.* Comparison of mannitol and methacholine to predict exercise-induced bronchoconstriction and a clinical diagnosis of asthma. *Respir Res* 2009; 10: 4.
- 53 Brown LL, Martin BL, Morris MJ. Airway hyperresponsiveness by methacholine challenge testing following negative exercise challenge. *J Asthma* 2004; 41: 553–558.
- 54 Holzer K, Anderson SD, Douglass J. Exercise in elite summer athletes: challenges for diagnosis. *J Allergy Clin Immunol* 2002; 110: 374–380.
- 55 Carlsen KH, Engh G, Mørk M, *et al.* Cold air inhalation and exercise-induced bronchoconstriction in relationship to methacholine bronchial responsiveness: different patterns in asthmatic children and children with other chronic lung diseases. *Respir Med* 1998; 92: 308–315.
- 56 Sin BA, Yildiz OA, Dursun AB, *et al.* Airway hyperresponsiveness: a comparative study of methacholine and exercise challenges in seasonal allergic rhinitis with or without asthma. *J Asthma* 2009; 46: 486–491.
- 57 Stensrud T, Mykland KV, Gabrielsen K, *et al.* Bronchial hyperresponsiveness in skiers: field test versus methacholine provocation? *Med Sci Sports Exerc* 2007; 39: 1681–1686.
- 58 Andregnette-Roscigno V, Fernández-Nieto M, Arochena L, *et al.* Methacholine is more sensitive than mannitol for evaluation of bronchial hyper-responsiveness in youth athletes with exercise-induced bronchoconstriction. *Pediatr Allergy Immunol* 2012; 23: 501–503.
- 59 Andregnette-Roscigno V, Fernández-Nieto M, Del Potro MG, *et al.* Methacholine is more sensitive than mannitol for evaluation of bronchial hyperresponsiveness in children with asthma. *J Allergy Clin Immunol* 2010; 126: 869–871.
- 60 Crapo RO, Casaburi R, Coates AL, *et al.* Guidelines for methacholine and exercise challenge testing - 1999. This official statement of the American Thoracic Society was adopted by the ATS Board of Directors, July 1999. *Am J Respir Crit Care Med* 2000; 161: 309–329.
- 61 Scheett TP, Nemet D, Stoppani J, *et al.* The effect of endurance-type exercise training on growth mediators and inflammatory cytokines in pre-pubertal and early pubertal males. *Pediatr Res* 2002; 52: 491–497.
- 62 Pedersen S, Hansen OR. Budesonide treatment of moderate and severe asthma in children: a dose-response study. *J Allergy Clin Immunol* 1995; 95: 29–33.
- 63 Clinical exercise testing with reference to lung diseases: indications, standardization and interpretation strategies. ERS Task Force on Standardization of Clinical Exercise Testing. *Eur Respir J* 1997; 10: 2662–2689.
- 64 Knöpfli BH, Bar-Or O, Araújo CG. Effect of ipratropium bromide on EIB in children depends on vagal activity. *Med Sci Sports Exerc* 2005; 37: 354–359.
- 65 Hancox RJ, Subbarao P, Kamada D, *et al.* β_2 -agonist tolerance and exercise-induced bronchospasm. *Am J Respir Crit Care Med* 2002; 165: 1068–1070.
- 66 Inman MD, O'Byrne PM. The effect of regular inhaled albuterol on exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 1996; 153: 65–69.
- 67 Kippelen P, Larsson J, Anderson SD, *et al.* Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. *Med Sci Sports Exerc* 2010; 42: 273–280.
- 68 Driessen JM, Nieland H, van der Palen JA, *et al.* Effects of a single dose inhaled corticosteroid on the dynamics of airway obstruction after exercise. *Pediatr Pulmonol* 2011; 46: 849–856.
- 69 Subbarao P, Duong M, Adelroth E, *et al.* Effect of ciclesonide dose and duration of therapy on exercise-induced bronchoconstriction in patients with asthma. *J Allergy Clin Immunol* 2006; 117: 1008–1013.
- 70 Haverkamp HC, Dempsey JA, Miller JD, *et al.* Repeat exercise normalizes the gas-exchange impairment induced by a previous exercise bout in asthmatic subjects. *J Appl Physiol* 2005; 99: 1843–1852.
- 71 Mickleborough TD, Lindley MR, Turner LA. Comparative effects of a high-intensity interval warm-up and salbutamol on the bronchoconstrictor response to exercise in asthmatic athletes. *Int J Sports Med* 2007; 28: 456–462.
- 72 Vianna EO, Boaventura LC, Terra-Filho J, *et al.* Morning-to-evening variation in exercise-induced bronchospasm. *J Allergy Clin Immunol* 2002; 110: 236–240.
- 73 Tecklenburg SL, Mickleborough TD, Fly AD, *et al.* Ascorbic acid supplementation attenuates exercise-induced bronchoconstriction in patients with asthma. *Respir Med* 2007; 101: 1770–1778.
- 74 Miller MR, Hankinson J, Brusasco V, *et al.* Standardisation of spirometry. *Eur Respir J* 2005; 26: 319–338.

- 75 Kallings LV, Emtner M, Bäcklund L. Exercise-induced bronchoconstriction in adults with asthma – comparison between running and cycling and between cycling at different air conditions. *Ups J Med Sci* 1999; 104: 191–198.
- 76 Hallstrand TS, Curtis JR, Koeppell TD, *et al.* Effectiveness of screening examinations to detect unrecognized exercise-induced bronchoconstriction. *J Pediatr* 2002; 141: 343–348.
- 77 Eliasson AH, Phillips YY, Rajagopal KR, *et al.* Sensitivity and specificity of bronchial provocation testing. An evaluation of four techniques in exercise-induced bronchospasm. *Chest* 1992; 102: 347–355.
- 78 Haby MM, Anderson SD, Peat JK, *et al.* An exercise challenge protocol for epidemiological studies of asthma in children: comparison with histamine challenge. *Eur Respir J* 1994; 7: 43–49.
- 79 Heaman DJ, Estes J. The free-running asthma screening test: an approach to screening for exercise-induced asthma in rural Alabama. *J Sch Health* 1997; 67: 83–88.
- 80 Helenius IJ, Tikkanen HO, Haahtela T. Occurrence of exercise induced bronchospasm in elite runners: dependence on atopy and exposure to cold air and pollen. *Br J Sports Med* 1998; 32: 125–129.
- 81 Rundell KW, Anderson SD, Spiering BA, *et al.* Field exercise vs laboratory eucapnic voluntary hyperventilation to identify airway hyperresponsiveness in elite cold weather athletes. *Chest* 2004; 125: 909–915.
- 82 Rundell KW, Wilber RL, Szmedra L, *et al.* Exercise-induced asthma screening of elite athletes: field versus laboratory exercise challenge. *Med Sci Sports Exerc* 2000; 32: 309–316.
- 83 Carlsen KH, Engh G, Mørk M. Exercise-induced bronchoconstriction depends on exercise load. *Respir Med* 2000; 94: 750–755.
- 84 Anderson SD, Brannan JD. Methods for “indirect” challenge tests including exercise, eucapnic voluntary hyperpnea, and hypertonic aerosols. *Clin Rev Allergy Immunol* 2003; 24: 27–54.
- 85 Villaran C, O’Neill SJ, Helbling A, *et al.* Montelukast versus salmeterol in patients with asthma and exercise-induced bronchoconstriction. Montelukast/Salmeterol Exercise Study Group. *J Allergy Clin Immunol* 1999; 104: 547–553.
- 86 Trümper C, Mäueler S, Vobejda C, *et al.* Heart rate-based protocols for exercise challenge testing do not ensure sufficient exercise intensity for inducing exercise-induced bronchial obstruction. *Br J Sports Med* 2009; 43: 429–431.
- 87 Hofstra WB, Sont JK, Sterk PJ, *et al.* Sample size estimation in studies monitoring exercise-induced bronchoconstriction in asthmatic children. *Thorax* 1997; 52: 739–741.
- 88 Leff JA, Busse WW, Pearlman D, *et al.* Montelukast, a leukotriene-receptor antagonist, for the treatment of mild asthma and exercise-induced bronchoconstriction. *N Engl J Med* 1998; 339: 147–152.
- 89 Anderson SD, Pearlman DS, Rundell KW, *et al.* Reproducibility of the airway response to an exercise protocol standardized for intensity, duration, and inspired air conditions, in subjects with symptoms suggestive of asthma. *Respir Res* 2010; 11: 120.
- 90 De Fuccio MB, Nery LE, Malaguti C, *et al.* Clinical role of rapid-incremental tests in the evaluation of exercise-induced bronchoconstriction. *Chest* 2005; 128: 2435–2442.
- 91 Rosenthal RR. Simplified eucapnic voluntary hyperventilation challenge. *J Allergy Clin Immunol* 1984; 73: 676–679.
- 92 Phillips YY, Jaeger JJ, Laube BL, *et al.* Eucapnic voluntary hyperventilation of compressed gas mixture. A simple system for bronchial challenge by respiratory heat loss. *Am Rev Respir Dis* 1985; 131: 31–35.
- 93 Argyros GJ, Roach JM, Hurwitz KM, *et al.* Eucapnic voluntary hyperventilation as a bronchoprovocation technique: development of a standardized dosing schedule in asthmatics. *Chest* 1996; 109: 1520–1524.
- 94 Spiering BA, Judelson DA, Rundell KW. An evaluation of standardizing target ventilation for eucapnic voluntary hyperventilation using FEV₁. *J Asthma* 2004; 41: 745–749.
- 95 Stadelmann K, Stensrud T, Carlsen KH. Respiratory symptoms and bronchial responsiveness in competitive swimmers. *Med Sci Sports Exerc* 2011; 43: 375–381.
- 96 Hurwitz KM, Argyros GJ, Roach JM, *et al.* Interpretation of eucapnic voluntary hyperventilation in the diagnosis of asthma. *Chest* 1995; 108: 1240–1245.
- 97 Mannix ET, Manfredi F, Farber MO. A comparison of two challenge tests for identifying exercise-induced bronchospasm in figure skaters. *Chest* 1999; 115: 649–653.
- 98 Mannix ET, Roberts MA, Dukes HJ, *et al.* Airways hyperresponsiveness in high school athletes. *J Asthma* 2004; 41: 567–574.
- 99 Mannix ET, Roberts M, Fagin DP, *et al.* The prevalence of airways hyperresponsiveness in members of an exercise training facility. *J Asthma* 2003; 40: 349–355.
- 100 Murray CS, Woodcock A, Langley SJ, *et al.* Secondary prevention of asthma by the use of Inhaled Fluticasone propionate in Wheezy Infants (IFWIN): double-blind, randomised, controlled study. *Lancet* 2006; 368: 754–762.
- 101 Pedersen L, Winther S, Backer V, *et al.* Airway responses to eucapnic hyperpnea, exercise, and methacholine in elite swimmers. *Med Sci Sports Exerc* 2008; 40: 1567–1572.
- 102 Dickinson JW, Whyte GP, McConnell AK, *et al.* Screening elite winter athletes for exercise induced asthma: a comparison of three challenge methods. *Br J Sports Med* 2006; 40: 179–182.
- 103 McFadden ER Jr, Nelson JA, Skowronski ME, *et al.* Thermally induced asthma and airway drying. *Am J Respir Crit Care Med* 1999; 160: 221–226.
- 104 Evans TM, Rundell KW, Beck KC, *et al.* Airway narrowing measured by spirometry and impulse oscillometry following room temperature and cold temperature exercise. *Chest* 2005; 128: 2412–2419.
- 105 Evans TM, Rundell KW, Beck KC, *et al.* Cold air inhalation does not affect the severity of EIB after exercise or eucapnic voluntary hyperventilation. *Med Sci Sports Exerc* 2005; 37: 544–549.
- 106 Sylvester KP, O’Connor BJ, Farebrother HM, *et al.* Cold air and exercise challenge – influence of minute ventilation. *J Asthma* 2007; 44: 143–147.
- 107 Nielsen KG, Bisgaard H. Hyperventilation with cold versus dry air in 2- to 5-year-old children with asthma. *Am J Respir Crit Care Med* 2005; 171: 238–241.
- 108 Koskela H, Tukiainen H. Facial cooling, but not nasal breathing of cold air, induces bronchoconstriction: a study in asthmatic and healthy subjects. *Eur Respir J* 1995; 8: 2088–2093.
- 109 Zeitoun M, Wilk B, Matsuzaka A, *et al.* Facial cooling enhances exercise-induced bronchoconstriction in asthmatic children. *Med Sci Sports Exerc* 2004; 36: 767–771.

- 110 de Benedictis FM, Canny GJ, MacLusky IB, *et al.* Comparison of airway reactivity induced by cold air and metacholine challenges in asthmatic children. *Pediatr Pulmonol* 1995; 19: 326–329.
- 111 Steinbrugger B, Eber E, Modl M, *et al.* A comparison of a single-step cold-dry air challenge and a routine histamine provocation for the assessment of bronchial responsiveness in children and adolescents. *Chest* 1995; 108: 741–745.
- 112 Schweitzer C, Vu LT, Nguyen YT, *et al.* Estimation of the bronchodilatory effect of deep inhalation after a free run in children. *Eur Respir J* 2006; 28: 89–95.
- 113 Slats AM, Janssen K, van Schadewijk A, *et al.* Bronchial inflammation and airway responses to deep inspiration in asthma and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2007; 176: 121–128.
- 114 Scichilone N, Permutt S, Toghias A. The lack of the bronchoprotective and not the bronchodilatory ability of deep inspiration is associated with airway hyperresponsiveness. *Am J Respir Crit Care Med* 2001; 163: 413–419.
- 115 Kapsali T, Permutt S, Laube B, *et al.* Potent bronchoprotective effect of deep inspiration and its absence in asthma. *J Appl Physiol* 2000; 89: 711–720.
- 116 Brudno DS, Wagner JM, Rupp NT. Length of postexercise assessment in the determination of exercise-induced bronchospasm. *Ann Allergy* 1994; 73: 227–231.
- 117 Backer V, Ulrik CS. Bronchial responsiveness to exercise in a random sample of 494 children and adolescents from Copenhagen. *Clin Exp Allergy* 1992; 22: 741–747.
- 118 Godfrey S, Springer C, Noviski N, *et al.* Exercise but not methacholine differentiates asthma from chronic lung disease in children. *Thorax* 1991; 46: 488–492.
- 119 Kattan M, Keens TG, Mellis CM, *et al.* The response to exercise in normal and asthmatic children. *J Pediatr* 1978; 92: 718–721.
- 120 Tilles SA. Exercise-induced respiratory symptoms: an epidemic among adolescents. *Ann Allergy Asthma Immunol* 2010; 104: 361–367.
- 121 Porsbjerg C, Sverrild A, Backer V. The usefulness of the mannitol challenge test for asthma. *Expert Rev Respir Med* 2013; 7: 655–663.
- 122 Choi IS, Chung SW, Koh YI, *et al.* Airway hyperresponsiveness to hypertonic saline as a predictive index of exercise-induced bronchoconstriction. *Korean J Intern Med* 2005; 20: 284–289.
- 123 Riedler J, Reade T, Dalton M, *et al.* Hypertonic saline challenge in an epidemiologic survey of asthma in children. *Am J Respir Crit Care Med* 1994; 150: 1632–1639.
- 124 Smith CM, Anderson SD. Inhalational challenge using hypertonic saline in asthmatic subjects: a comparison with responses to hyperpnoea, methacholine and water. *Eur Respir J* 1990; 3: 144–151.
- 125 Porsbjerg C, Brannan JD, Anderson SD, *et al.* Relationship between airway responsiveness to mannitol and to methacholine and markers of airway inflammation, peak flow variability and quality of life in asthma patients. *Clin Exp Allergy* 2008; 38: 43–50.
- 126 Gibson PG, Saltos N, Borgas T. Airway mast cells and eosinophils correlate with clinical severity and airway hyperresponsiveness in corticosteroid-treated asthma. *J Allergy Clin Immunol* 2000; 105: 752–759.
- 127 Mai XM, Böttcher MF, Bruhammar M, *et al.* Urinary inflammatory mediators and inhalation of hypertonic saline in children. *Allergy* 2005; 60: 60–64.
- 128 Rodwell LT, Anderson SD, Seale JP. Inhaled clemastine, an H1 antihistamine inhibits airway narrowing caused by aerosols of non-isotonic saline. *Eur Respir J* 1991; 4: 1126–1134.
- 129 Umeno E, McDonald DM, Nadel JA. Hypertonic saline increases vascular permeability in the rat trachea by producing neurogenic inflammation. *J Clin Invest* 1990; 85: 1905–1908.
- 130 Anderson SD, Brannan J, Spring J, *et al.* A new method for bronchial-provocation testing in asthmatic subjects using a dry powder of mannitol. *Am J Respir Crit Care Med* 1997; 156: 758–765.
- 131 Brannan JD, Koskela H, Anderson SD, *et al.* Responsiveness to mannitol in asthmatic subjects with exercise- and hyperventilation-induced asthma. *Am J Respir Crit Care Med* 1998; 158: 1120–1126.
- 132 Muñoz PA, Gómez FP, Manrique HA, *et al.* Pulmonary gas exchange response to exercise- and mannitol-induced bronchoconstriction in mild asthma. *J Appl Physiol* 2008; 105: 1477–1485.
- 133 Kersten ET, Driessen JM, van der Berg JD, *et al.* Mannitol and exercise challenge tests in asthmatic children. *Pediatr Pulmonol* 2009; 44: 655–661.
- 134 Sverrild A, Bergqvist A, Baines KJ, *et al.* Airway responsiveness to mannitol in asthma is associated with chymase-positive mast cells and eosinophilic airway inflammation. *Clin Exp Allergy* 2016; 46: 288–297.
- 135 Brannan JD, Gulliksson M, Anderson SD, *et al.* Inhibition of mast cell PGD2 release protects against mannitol-induced airway narrowing. *Eur Respir J* 2006; 27: 944–950.
- 136 Avital A, Godfrey S, Springer C. Exercise, methacholine, and adenosine 5'-monophosphate challenges in children with asthma: relation to severity of the disease. *Pediatr Pulmonol* 2000; 30: 207–214.
- 137 Perzanowski MS, Yoo Y. Exhaled nitric oxide and airway hyperresponsiveness to adenosine 5'-monophosphate and methacholine in children with asthma. *Int Arch Allergy Immunol* 2015; 166: 107–113.
- 138 van den Berge M, Kerstjens HA, Meijer RJ, *et al.* Corticosteroid-induced improvement in the PC20 of adenosine monophosphate is more closely associated with reduction in airway inflammation than improvement in the PC20 of methacholine. *Am J Respir Crit Care Med* 2001; 164: 1127–1132.
- 139 Hua X, Chason KD, Fredholm BB, *et al.* Adenosine induces airway hyperresponsiveness through activation of A3 receptors on mast cells. *J Allergy Clin Immunol* 2008; 122: 107–113.
- 140 Saetta M, Di Stefano A, Turato G, *et al.* Fatal asthma attack during an inhalation challenge with ultrasonically nebulized distilled water. *J Allergy Clin Immunol* 1995; 95: 1285–1287.
- 141 Smith CM, Anderson SD. Inhalation provocation tests using nonisotonic aerosols. *J Allergy Clin Immunol* 1989; 84: 781–790.
- 142 Perpiñá Tordera M, García Río F, Álvarez Gutierrez FJ, *et al.* Guidelines for the study of nonspecific bronchial hyperresponsiveness in asthma. Spanish Society of Pulmonology and Thoracic Surgery (SEPAR). *Arch Bronconeumol* 2013; 49: 432–446.
- 143 Anderson SD. Indirect challenge tests: airway hyperresponsiveness in asthma: its measurement and clinical significance. *Chest* 2010; 138: Suppl. 2, 25S–30S.
- 144 de Meer G, Marks GB, de Jongste JC, *et al.* Airway responsiveness to hypertonic saline: dose-response slope or PD15? *Eur Respir J* 2005; 25: 153–158.

- 145 du Toit JI, Anderson SD, Jenkins CR, *et al.* Airway responsiveness in asthma: bronchial challenge with histamine and 4.5% sodium chloride before and after budesonide. *Allergy Asthma Proc* 1997; 18: 7–14.
- 146 Rodwell LT, Anderson SD, Seale JP. Inhaled steroids modify bronchial responses to hyperosmolar saline. *Eur Respir J* 1992; 5: 953–962.
- 147 Gibson PG, Wlodarczyk JW, Hensley MJ, *et al.* Epidemiological association of airway inflammation with asthma symptoms and airway hyperresponsiveness in childhood. *Am J Respir Crit Care Med* 1998; 158: 36–41.
- 148 Smith CM, Anderson SD. A comparison between the airway response to isocapnic hyperventilation and hypertonic saline in subjects with asthma. *Eur Respir J* 1989; 2: 36–43.
- 149 Leuppi JD, Anderson SD, Brannan JD, *et al.* Questionnaire responses that predict airway response to hypertonic saline. *Respiration* 2005; 72: 52–60.
- 150 Riedler J, Gamper A, Eder W, *et al.* Prevalence of bronchial hyperresponsiveness to 4.5% saline and its relation to asthma and allergy symptoms in Austrian children. *Eur Respir J* 1998; 11: 355–360.
- 151 Araki H, Sly PD. Inhalation of hypertonic saline as a bronchial challenge in children with mild asthma and normal children. *J Allergy Clin Immunol* 1989; 84: 99–107.
- 152 Gibson PG, Saltos N, Fakes K. Acute anti-inflammatory effects of inhaled budesonide in asthma: a randomized controlled trial. *Am J Respir Crit Care Med* 2001; 163: 32–36.
- 153 Anderson SD, du Toit JI, Rodwell LT, *et al.* Acute effect of sodium cromoglycate on airway narrowing induced by 4.5 percent saline aerosol. Outcome before and during treatment with aerosol corticosteroids in patients with asthma. *Chest* 1994; 105: 673–680.
- 154 Anderson SD. “Indirect” challenges from science to clinical practice. *Eur Clin Respir J* 2016; 3: 31096.
- 155 Brannan JD, Anderson SD, Perry CP, *et al.* The safety and efficacy of inhaled dry powder mannitol as a bronchial provocation test for airway hyperresponsiveness: a phase 3 comparison study with hypertonic (4.5%) saline. *Respir Res* 2005; 6: 144.
- 156 O’Connor G, Sparrow D, Taylor D, *et al.* Analysis of dose-response curves to methacholine. An approach suitable for population studies. *Am Rev Respir Dis* 1987; 136: 1412–1417.
- 157 Abramson MJ, Saunders NA, Hensley MJ. Analysis of bronchial reactivity in epidemiological studies. *Thorax* 1990; 45: 924–929.
- 158 Sverrild A, Porsbjerg C, Thomsen SF, *et al.* Airway hyperresponsiveness to mannitol and methacholine and exhaled nitric oxide: a random-sample population study. *J Allergy Clin Immunol* 2010; 126: 952–958.
- 159 Sverrild A, Porsbjerg C, Thomsen SF, *et al.* Diagnostic properties of inhaled mannitol in the diagnosis of asthma: a population study. *J Allergy Clin Immunol* 2009; 124: 928–932.
- 160 Kononowa N, Michel S, Miedinger D, *et al.* Effects of add-on montelukast on airway hyperresponsiveness in patients with well-controlled asthma – a pilot study. *J Drug Assess* 2013; 2: 49–57.
- 161 Brannan JD, Perry CP, Anderson SD. Mannitol test results in asthmatic adults receiving inhaled corticosteroids. *J Allergy Clin Immunol* 2013; 131: 906–907.
- 162 Koskela HO, Hyvärinen L, Brannan JD, *et al.* Sensitivity and validity of three bronchial provocation tests to demonstrate the effect of inhaled corticosteroids in asthma. *Chest* 2003; 124: 1341–1349.
- 163 Brannan JD, Koskela H, Anderson SD, *et al.* Budesonide reduces sensitivity and reactivity to inhaled mannitol in asthmatic subjects. *Respirology* 2002; 7: 37–44.
- 164 Lipworth BJ, Short PM, Williamson PA, *et al.* A randomized primary care trial of steroid titration against mannitol in persistent asthma: STAMINA trial. *Chest* 2012; 141: 607–615.
- 165 Clearie KL, Jackson CM, Fardon TC, *et al.* Supervised step-down of inhaled corticosteroids in the community – an observational study. *Respir Med* 2011; 105: 558–565.
- 166 Leuppi JD, Salome CM, Jenkins CR, *et al.* Predictive markers of asthma exacerbation during stepwise dose reduction of inhaled corticosteroids. *Am J Respir Crit Care Med* 2001; 163: 406–412.
- 167 Miedinger D, Mosimann N, Meier R, *et al.* Asthma tests in the assessment of military conscripts. *Clin Exp Allergy* 2010; 40: 224–231.
- 168 van den Berge M, Polosa R, Kerstjens HA, *et al.* The role of endogenous and exogenous AMP in asthma and chronic obstructive pulmonary disease. *J Allergy Clin Immunol* 2004; 114: 737–746.
- 169 De Meer G, Heederik DJ, Brunekreef B, *et al.* Repeatability of bronchial hyperresponsiveness to adenosine 5′-monophosphate (AMP) by a short dosimeter protocol. *Thorax* 2001; 56: 362–365.
- 170 Prieto L, Reig I, Rojas R, *et al.* The effect of challenge method on sensitivity and reactivity to adenosine 5′-monophosphate in subjects with suspected asthma. *Chest* 2006; 130: 1448–1453.
- 171 Lexmond AJ, Hagedoorn P, Frijlink HW, *et al.* Challenging the two-minute tidal breathing challenge test. *J Aerosol Med Pulm Drug Deliv* 2013; 26: 380–386.
- 172 Lexmond AJ, van der Wiel E, Hagedoorn P, *et al.* Adenosine dry powder inhalation for bronchial challenge testing, part 2: proof of concept in asthmatic subjects. *Eur J Pharm Biopharm* 2014; 88: 148–152.
- 173 Lexmond AJ, Hagedoorn P, van der Wiel E, *et al.* Adenosine dry powder inhalation for bronchial challenge testing, part 1: inhaler and formulation development and *in vitro* performance testing. *Eur J Pharm Biopharm* 2014; 86: 105–114.
- 174 Van Schoor J, Pauwels R, Joos G. Indirect bronchial hyper-responsiveness: the coming of age of a specific group of bronchial challenges. *Clin Exp Allergy* 2005; 35: 250–261.
- 175 Choi SH, Kim DK, Yu J, *et al.* Bronchial responsiveness to methacholine and adenosine 5′-monophosphate in young children with asthma: their relationship with blood eosinophils and serum eosinophil cationic protein. *Allergy* 2007; 62: 1119–1124.
- 176 Suh DI, Lee JK, Kim CK, *et al.* Bronchial hyperresponsiveness to methacholine/AMP and the bronchodilator response in asthmatic children. *Eur Respir J* 2011; 37: 800–805.
- 177 Park GM, Han HW, Kim JY, *et al.* Association of symptom control with changes in lung function, bronchial hyperresponsiveness, and exhaled nitric oxide after inhaled corticosteroid treatment in children with asthma. *Allergol Int* 2016; 65: 439–443.
- 178 Spicuzza L, Scuderi V, Morjaria JB, *et al.* Airway responsiveness to adenosine after a single dose of fluticasone propionate discriminates asthma from COPD. *Pulm Pharmacol Ther* 2014; 27: 70–75.
- 179 van den Berge M, Luijk B, Bareille P, *et al.* Prolonged protection of the new inhaled corticosteroid fluticasone furoate against AMP hyperresponsiveness in patients with asthma. *Allergy* 2010; 65: 1531–1535.

- 180 Mastruzzo C, Contrafatto MR, Crimi C, *et al.* Acute additive effect of montelukast and beclomethasone on AMP induced bronchoconstriction. *Respir Med* 2010; 104: 1417–1424.
- 181 Derom E, Van De Velde V, Marissens S, *et al.* Effects of inhaled ciclesonide and fluticasone propionate on cortisol secretion and airway responsiveness to adenosine 5′monophosphate in asthmatic patients. *Pulm Pharmacol Ther* 2005; 18: 328–336.
- 182 Gauvreau GM, Watson RM, Rerecich TJ, *et al.* Repeatability of allergen-induced airway inflammation. *J Allergy Clin Immunol* 1999; 104: 66–71.
- 183 Cockcroft DW, Murdock KY, Kirby J, *et al.* Prediction of airway responsiveness to allergen from skin sensitivity to allergen and airway responsiveness to histamine. *Am Rev Respir Dis* 1987; 135: 264–267.
- 184 Chen R, Smith SG, Salter B, *et al.* Allergen-induced increases in sputum levels of group 2 innate lymphoid cells in subjects with asthma. *Am J Respir Crit Care Med* 2017; 196: 700–712.
- 185 Lee YG, Jeong JJ, Nyenhuis S, *et al.* Recruited alveolar macrophages, in response to airway epithelial-derived monocyte chemoattractant protein 1/CCL2, regulate airway inflammation and remodeling in allergic asthma. *Am J Respir Cell Mol Biol* 2015; 52: 772–784.
- 186 O’Byrne PM. Allergen-induced airway inflammation and its therapeutic intervention. *Allergy Asthma Immunol Res* 2009; 1: 3–9.